SAMPLE PREP COLUMNS DESIGNED SPECIFICALLY FOR THE RACING INDUSTRY
XtrackT® DAU is a series of reproducible copolymerically bonded silicas created especially for the screening and confirmation of drugs in dog and horse urine.

The use of new and powerful drugs in sports requires new techniques for the clean extraction of very low levels of compounds. XtrackT® offers a simple, easy solution to this extraction problem, whether it's a comprehensive screen or low level quantitation by either GC/MS or LC/MS.

Through original research on the concept and use of copolymeric bonding of silicas for sample preparation, UCT, Inc. has pioneered this generation of hybrid extraction sorbents.

XtrackT® DAU has both hydrophobic and ion exchange functionalities, thus providing several primary retention mechanisms. The copolymers are used to enhance and improve mixed mechanisms which have been known to exist for sometime. XtrackT® utilizes several different chemical characteristics of compounds to produce very clean extracts.

The recovery of drugs is at least equivalent to and in most cases significantly better than recovery by liquid-liquid extraction. XtrackT® extracts both free and glucuronide bound drugs.

A single column extraction provides broad coverage of drugs, separating extracts into acidic/neutral, steroid and basic fractions. It produces cleaner extracts and eliminates the need for special liquid-liquid extraction procedures for different drug classes. The columns are designed to give uniform flow even with the most viscous of samples.

A SCREENING PROCEDURE FOR ACIDIC, NEUTRAL AND BASIC DOPING AGENTS FROM HUMAN, EQUINE AND CANINE URINE USING XtrackT® EXTRACTION COLUMNS

1. Hydrolyze conjugates
2. Adjust sample pH to 6.0
3. Condition column
4. Apply sample to column
5. Wash
6. Dry column
7. Elute acidic and neutral drugs
8. Elute steroids
9. Wash column
10. Elute basic drugs
11. Evaporate and reconstitute
3-HYDROXY LIDOCAINE, 4-HYDROXY GUANABENZ, 4-HYDROXY MEPIVICANE, 4-HYDROXY XYLAZINE, DETOMIDINE, AND O-DESMETHYL TRAMADOL IN EQUINE URINE BY LC/MS

200 mg XtrackT® DAU Extraction Column - Part #: XRDAH206
Select pH Buffer Pouches 100mM Phosphate pH 6.0 - Part #: SPHPHO6001-10

1. Prepare Sample:
To 1 mL of 100 mM phosphate buffer (pH= 6) add 2 mL of Urine
Add Internal standards. Add 3 mL of 100 mM phosphate buffer
Mix/ vortex
Centrifuge as appropriate

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH\textsubscript{3}OH
1 x 3 mL D.I. H\textsubscript{2}O
1 x 3 mL 100 mM phosphate buffer (pH= 6)
NOTE: Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 to 2 mL/ minute

4. Wash Column:
1 x 3 mL D.I. H\textsubscript{2}O
1 x 3 mL CH\textsubscript{3}OH/ 2% glacial acetic acid
Dry column (5 minutes at full vacuum or pressure)

5. Elute:
1 x 3 mL DCM/IPA/ NH\textsubscript{4}OH (78/20/2)
Collect the eluate at 1-2 mL minute (or gravity)

6. Dry Eluate:
Evaporate to dryness at < 40°C

7. Analysis:
Inject 10 μL sample

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<thead>
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<th>Compound</th>
<th>RT</th>
<th>Precursor Ion</th>
<th>Product Ion</th>
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<td>O-Desmethyl Tramadol</td>
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<td>4-Hydroxy Xylazine</td>
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<td>4-Hydroxy Guanabenz</td>
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<tr>
<td>Detomidine</td>
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<td>188.1</td>
<td>81.0</td>
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</table>
1. Prepare Sample:
To 1 mL of 100 mM phosphate buffer (pH 6) add 1 mL of Urine
Add Internal standards Add 3 mL of 100 mM phosphate buffer
Mix/ vortex
Centrifuge as appropriate

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH₃OH
1 x 3 mL D.I. H₂O
1 x 3 mL 100 mM phosphate buffer (pH 6)
NOTE: Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 to 2 mL/ minute

4. Wash Column:
1 x 3 mL D.I. H₂O
1 x 3 mL CH₃OH
Dry column (5 minutes at full vacuum or pressure)

5. Elute Clenbuterol / Salbutamol:
1 x 3 mL CH₃OH containing 4% NH₄OH
Collect the eluate at 1-2 mL minute (or gravity)

6. Dry Eluate:
Evaporate to dryness at < 40°C

7. Derivatize:
Add 50 μL Ethyl Acetate
Add 50 μL BSTFA w/1% TMCS
Heat at 70 °C for 30 minutes
Cool to room temperature
NOTE: Do not evaporate this solution

8. Analysis:
Inject 1 to 2 μL onto gas chromatograph

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**CLENBUTEROL AND SALBUTAMOL IN EQUINE URINE FOR GC/MS CONFIRMATIONS**

200 mg XtrackT® DAU Extraction Column - Part #: XRDASH206
Select pH Buffer Pouches 100mM Phosphate pH 6.0 - Part #: SPHPH0601-10
BSTFA w/1% TMCS - Part #: SBSTFA-1-1

Pre-measured salts for sample preparation

---

**Select pH Buffer Pouches**

---

**Compound** | **Primary Ion** | **Secondary Ion** | **Tertiary Ion**
--- | --- | --- | ---
Clenbuterol-TMS | 86 | 262 | 243
Clenbuterol-D3-TMS | 95 | 262 | 243
Salbutamol-TMS | 369 | 86 | 207
Salbutamol-D3-TMS | 372 | 86 | 210

*Quantitation Ion
†Suggested internal standard for GC/MS
COCAINE AND METABOLITES IN BLOOD, PLASMA/SERUM, URINE AND TISSUE FOR GC/MS CONFIRMATIONS

1. Prepare Sample:
To 1 mL of 100 mM phosphate buffer (pH 6.0) add internal standards. Add 2 mL of blood, plasma/serum, urine or 1 g (1:4) tissue homogenate. Mix/vortex. Add 2 mL of 100 mM phosphate buffer (pH 6). Mix/vortex.

Sample pH should be 6.0 ± 0.5
Adjust pH accordingly with 100 mM monobasic or dibasic sodium phosphate
Centrifuge as appropriate

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH₃OH
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM phosphate buffer (pH 6.0)
NOTE: Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 to 2 mL/minute

4. Wash Column:
1 x 3 mL D.I. H₂O
1 x 2 mL 100 mM HCl
1 x 3 mL CH₃OH
Dry column (5 minutes at full vacuum or pressure)

5. Elute Cocaine and Benzoylecgonine:
1 x 3 mL Methylene Chloride/Isopropanol/
Ammonium Hydroxide (78:20:2)
Collect eluate at 1 to 2 mL/minute
NOTE: Prepare elution solvent daily. Add IPA/NH₄OH, mix, then add CH₂Cl₂ (pH 11-12)

6. Dry Eluate:
Evaporate to dryness at < 40°C

7. Derivatize:
Add 50 μL ethyl acetate and 50 μL BSTFA w/1% TMCS
Overlay with Nitrogen and cap. Mix/vortex
React 20 minutes at 70°C
Remove from heat source to cool
NOTE: Do not evaporate BSTFA solution

8. Quantitate:
Inject 1 to 2 μL onto gas chromatograph

COCAINE AND METABOLITES IN BLOOD, PLASMA/SERUM, URINE AND TISSUE FOR GC/MS CONFIRMATIONS

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*Quantitation Ion
†Suggested internal standard for GC/MS
BARBITURATES IN URINE FOR GC/MS CONFIRMATIONS

1. Prepare Sample:
To 2 mL of urine add internal standard(s) and 1 mL of 100 mM phosphate buffer (pH = 5.0)
Mix/vortex.
Sample pH should be 5.0 ± 0.5
Adjust pH accordingly with 100 mM monobasic or dibasic sodium phosphate

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH₃OH
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM phosphate buffer (pH = 5.0)
NOTE: Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 to 2 mL/ minute

4. Wash Column:
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM acetic acid
Dry column (5 minutes at full vacuum or pressure)
1 x 2 mL hexane

5. Elute Barbituates:
1 x 3 mL hexane/ethyl acetate (50:50);
Collect eluate at 1 to 2 mL / minute

6. Dry Eluate:
Evaporate to dryness at < 40°C
Reconstitute with 100 μL ethyl acetate
OPTIONAL DERIVATIZATION
Add 25-50 μL of 0.2 M TMPAH
Reaction occurs in injection port
Inject 1 to 2 μL onto gas chromatograph

7. Quantitative
Add 50 μL of both Ethyl Acetate and BSTFA

Other Barbituates that can be extracted using this method

Underivatized

<table>
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*Quantitation Ion

Derivatized

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*Quantitation Ion
†Suggested internal standard for GC/MS
Underivatized

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<td>Hexobarbital†</td>
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<td>157</td>
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*Quantitation Ion
†Suggested internal standard for GC/MS
1. Prepare Sample - Enzymatic and Base Hyd Denzymatic and Base Hydrolysis of Glucuronides:

To 1 mL of urine add internal standard(s) and 50 μL of Beta Glucuronidase solution (*Haliotis rufescens*), add 2 mL of 1 M Acetate buffer pH = 5. Mix and incubate at 65 °C for 3 hours. Cool to room temperature.

Add 100 of 10 M NaOH. Mix/vortex. Hydrolyze for 20 minutes at 60°C. Cool before proceeding. Adjust sample pH to 3.0 with approx. 1.0 mL of glacial acetic acid. Check pH to insure that the pH value is ~ 3.0. Sample pH should be 6.0 ± 0.5. Adjust pH accordingly with 100 mM monobasic or dibasic sodium phosphate. Centrifuge as appropriate.

2. Condition XtrackT® DAU Extraction Column:

1 x 3 mL CH3OH
1 x 3 mL D.I. H2O
1 x 1 mL Acetate buffer (pH= 3.0)

**NOTE:** Aspirate at full vacuum or pressure.

3. Apply Sample:
Load at 1 to 2 mL/minute.

4. Wash Column:

1 x 2 mL D.I. H2O
1 x 2 mL 100 mM HCl/acetonitrile (95:5)
Dry column (5-10 minutes at full vacuum or pressure)
1 x 200 mL hexane; Aspirate. (Additional step to remove any residual moisture)

5. Elute Cannabinoids:

1 x 3 mL hexane/ethyl acetate/ glacial acetic acid (49:49:2)
Collect eluate at 1 to 2 mL/minute.

**NOTE:** Before proceeding, insure there are no water droplets at the bottom of the collection tube. This may increase drying time and decrease BSTFA derivatizing efficiency.

6. Dry Eluate:
Evaporate to dryness at < 40°C

7. Derivatize:
Add 50 μL ethyl acetate and 50 μL BSTFA w/1% TMCS.
Mix/vortex.
React 20 minutes at 70°C.
Remove from heat source to cool.

**NOTE:** Do not evaporate BSTFA. Inject 1 to 2 μL onto gas chromatograph.

---

**Compound** | **Primary Ion** | **Secondary Ion** | **Tertiary Ion**
--- | --- | --- | ---
THC-TMS | 371 | 343 | 366
THC-D3-TMS† | 374 | 346 | 889
THC-OH-TMS | 371 | 459 | 474
THC-OH-D3-TMS† | 374 | 462 | 471
THC-COOH-TMS | 371 | 473 | 488
THC-COOH-D3-TMS† | 374 | 476 | 491

*Quantitation Ion
†Suggested internal standard for GC/MS
1. Prepare Sample - β-Glucuronidase Hydrolysis:
To 2 mL of urine add internal standard(s) and 1 mL of β-glucuronidase solution.
β-glucuronidase solution contains: 5,000 F units/mL *Haliotis rufescens* in 100 mM acetate buffer (pH=5.0).
Mix/vortex.
Hydrolyze for 3 hours at 65°C.
Centrifuge for 10 minutes at 2000 rpm and discard pellet.

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH₂OH
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM phosphate buffer (pH= 6.0)
**NOTE:** Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 mL/ minute

4. Wash Column:
1 x 2 mL D.I. H₂O
1 x 2 mL 20% acetonitrile in 100 mM phosphate buffer (pH= 6.0)
Dry column (5 minutes at full vacuum or pressure)
1 x 2 mL hexane

5. Elute Benzodiazepines:
1 x 5 mL ethyl acetate containing 4% ammonium hydroxide
collect eluate at 1 to 2 mL/minute

6. Dry Eluate:
Evaporate to dryness at < 40°C

7. Derivatize:
Add 50 μL ethyl acetate and 50 μL BSTFA w/1% TMCS
Overlay with Nitrogen and cap. Mix/vortex
React 20 minutes at 70°C. Remove from heat source to cool
**NOTE:** Do not evaporate BSTFA solution

8. Quantitate:
Inject 1 to 2 μL onto gas chromatograph

---

**Compound** | **Primary Ion** | **Secondary Ion** | **Tertiary Ion**
--- | --- | --- | ---
1. Diazepam | 256 | 283 | 221
2. Nordazepam TBDMS | 327 | 383 | 369
3. Midazepam | 310 | 325 | 297
4. Oxazepam - 2TBDMS | 457 | 513 | 383
Oxazepam - D5 2TBDMS† | 462 | 519 | —
5. Temazepam | 357 | 283 | 385
6. 7-aminoclonazepam TBDMS | 342 | 399 | 328
7. Lorazepam 2TBDMS | 491 | 513 | 533
8. Clonazepam | 372 | 326 | 429
9. Alprazolam | 279 | 204 | 308
Alprazolam - D5† | 284 | 313 | —
10. Alphahydroxy alprazolam TBDMS | 381 | 423 | 346

*Quantitation Ion
†Suggested internal standard for GC/MS
CARISOPRODOL/MEPROBAMATE IN URINE FOR GC/MS CONFIRMATIONS

200 mg XtrackT® DAU Extraction Column - Part #: XRDAH206
Select pH Buffer Pouches 100mM Phosphate pH 6.0 - Part #: SPHPH0601-10
BSTFA w/1% TMCS - Part #: SBSTFA-1-1

1. Prepare Sample:
To 2 mL of urine add internal standard(s) and 1 mL of 100 mM phosphate buffer (pH = 6)
Mix/vortex
Sample pH should be 6.0 ± 0.5
Adjust pH accordingly with 100 mM monobasic or dibasic sodium phosphate
Centrifuge at 3000 RPM for 10 minutes

2. Condition XtrackT® DAU Extraction Column:
1 x 3 mL CH₃OH
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM phosphate buffer (pH = 6)
NOTE: Aspirate at full vacuum or pressure

3. Apply Sample:
Load at 1 to 2 mL/ minute

4. Wash Column:
1 x 3 mL D.I. H₂O
1 x 1 mL 100 mM acetic acid
Dry column (5 minutes at full vacuum or pressure)
1 x 2 mL hexane

5. Elute Barbituates:
1 x 3 mL hexane/ethyl acetate (50:50); Collect eluate at 1 to 2 mL / minute

6. Dry Eluate:
Evaporate to dryness at < 40°C
Reconstitute with 100 μL ethyl acetate

7. Derivatize:
Add 50μL ethyl acetate and 50μL BSTFA w/1% TMCS
Mix/vortex
React 20 minutes at 70°C
Remove from heat source to cool
NOTE: Do not evaporate BSTFA

8. Quantitative:
Inject 1 to 2 μL onto gas chromatograph

Elution Profile – (1) Carisoprodol, (2) Meprobamate, (3) Hexobarbital (Internal Standard)

<table>
<thead>
<tr>
<th>Compound</th>
<th>Primary Ion</th>
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</thead>
<tbody>
<tr>
<td>Carisoprodol</td>
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</tr>
<tr>
<td>Meprobamate</td>
<td>157</td>
</tr>
<tr>
<td>Hexobarbital</td>
<td>236</td>
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</tbody>
</table>
BUPRENORPHINE AND NORBUPRENORPHINE IN EQUINE URINE FOR GC/MS CONFIRMATIONS

1. Prepare Sample:
   To 1 mL of 100 mM Acetate buffer (pH= 5) add internal standard.
   Mix/ vortex and add 1 mL of Equine Urine.
   Add 2 mL of 100 mM Acetate buffer (pH= 5) and mix/ vortex

   Sample pH should be 6.0 ± 0.5.
   Adjust pH accordingly with 100 mM monobasic or dibasic sodium phosphate.
   Centrifuge as appropriate.

   Enzyme Hydrolysis of Glucuronides.
   To 1 mL of 100 mM Acetate buffer add internal standard.
   Add 1 mL of Equine Urine. Mix/ vortex.
   Add 2 mL of 100 mM Acetate buffer (pH= 5).
   Hydrolyze with Helix Pomatia (5,000 units/mL), heat for 3 hours at 60°C Cool before proceeding.

2. Condition XtrackT® DAU Extraction Column:
   1 x 3 mL CH₃OH
   1 x 3 mL D.I. H₂O
   1 x 1 mL 100 mM Acetate buffer (pH= 5.0)
   **NOTE:** Aspirate at < 3 Inches Hg to prevent sorbent drying

3. Apply Sample:
   Load at 1 to 2 mL/ minute

4. Wash Column:
   1 x 2 mL D.I. H₂O
   1 x 3 mL 100 mM acetate buffer (pH=5.0)
   1 x 3 mL Methanol
   Dry column (5-10 minutes at full vacuum or pressure

5. Elute Buprenorphine / Norbuprenorphine:
   1 x 3 mL methylene chloride / iso-propano / ammonium hydroxide (78/20/12). (Make elution solvent fresh)
   Collect eluate at 1 to 2 mL/minute
   **NOTE:** Before proceeding, insure there are no water droplets
   at the bottom of the collection tube. This may increase drying time and decrease BSTFA derivatizing efficiency

6. Dry Eluate:
   Evaporate to dryness at < 40°C

7. Derivatize:
   Add 50 μL ethyl acetate and 50 μL BSTFA w/1% TMCS
   React 20 minutes at 70°C
   Remove from heat source to cool
   **NOTE:** Do not evaporate BSTFA

8. Quantitative:
   Inject 1 to 2 μL onto gas chromatograph/mass spectrometer

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**Compound** | **Primary Ion** | **Secondary Ion** | **Tertiary Ion**
---|---|---|---
Buprenorphine-TMS | 452 | 467 | 487
Buprenorphine-D₄-TMS† | 455 | 470 | 489
Norbuprenorphine-TMS | 468 | 500 | 510
Norbuprenorphine-D₅-TMS† | 503 | 525 | 542

*Quantitation Ion
†Suggested internal standard for GC/MS
PRICES AND TERMS
Our prices are subject to change without notice. The price in effect when we receive your order will apply. All prices are in US Dollars and are F.O.B. Lewistown, PA 17044. Terms of payment are net 30 days.

MINIMUM ORDERS
We welcome all orders, therefore, we do not have a minimum order requirement. When ordering, please include your purchase order number, complete “Ship To” and “Bill To” address, catalog number, quantity, and description of product(s). Also include your name and a phone number where you can be reached should we have any questions concerning your order.

SHIPMENTS
Normal processing is within 24 hours after receipt of an order. Unless special shipping requests have been made, our trained staff will send all orders by UPS Ground service. The appropriate shipping charges (freight & insurance costs) will be added to the invoice, unless otherwise instructed by the customer.

SPECIAL PRICING
We offer special pricing for volume purchases and standing orders. These discounts apply to bonded phase extraction column purchases only. Please call a sales representative for more information on special pricing qualifications.

RETURN POLICY
Our Quality Manager will handle all returns. Before returning merchandise, please call to obtain a return authorization number from the quality manager. We will need to know the reason for the return, date of purchase, purchase order number and invoice number in order to issue a return authorization number. Return merchandise must be received before a credit can be issued. Returns will not be accepted after 90 days. A restocking fee of 25% of the price paid, or a minimum of $25.00 (whichever is greater) will be charged on all returns.

WARRANTY
All products manufactured by UCT are guaranteed against defects in materials and workmanship for a period of 90 days after shipment. UCT will replace any items that prove to be defective during this time period. The exclusive remedy requires the end user to first advise UCT of the defective product by phone or in writing and must include order number, the lot number and the shipping date. To initiate this action, photographs of the product, including packaging and labeling of the containers, must be submitted to the UCT Representative for approval. With approval a return authorization can be initiated, and must be received within 30 days. Once the materials arrive at UCT a further inspection of the materials must be completed and accepted by our Quality Manager prior to further action of credits or replacement. UCT’s total liability is limited to the replacement cost of UCT products. This warranty does not apply to damage resulting from misuse.

Select Biobibliography of XtrackT® publications from the racing industry

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Web: www.unitedchem.com